Implementation of a Multiplex Molecular Gastrointestinal Pathogen Panel: The Memorial Healthcare System Experience Rodney C. Arcenas, PhD, D(ABMM)^{1,2}, <u>M. Liliana Bedoya, MBA, BS, MT(AMT)², Frederick L. Kiechle, MD, PhD^{1,2}</u> ¹Pathology Consultants of South Broward, Hollywood, FL and ²Memorial Healthcare System, Hollywood, FL

Abstract (Revised and Updated)

Background: Infectious gastroenteritis can be a serious medical and economic burden to healthcare institutions. Many of the clinical symptoms overlap making it difficult for clinicians to make an accurate diagnosis without the use of laboratory testing. Traditional microbiological methods for stool pathogen testing have long been a mainstay in the clinical laboratory which can be time consuming, labor intensive, and involve multiple tests if bacterial, viral, and parasitic pathogens are being investigated. There is a shift towards syndromic testing involving multiplex molecular methods. This abstract describes the experience at the Memorial Healthcare System of implementing the Luminex xTAG[®] GastroPathogen Panel (GPP). Methods: The GPP detects 13 (bacterial, viral, and parasitic) pathogens. This assay was performed as per package insert instructions utilizing the NucliSENS[®] easyMAG (bioMerieux) for stool extractions. Samples were obtained from Zeptometrix NATrol[™] (11), Luminex (119), and left-over stool samples (105) from our Microbiology laboratory that were processed for culture and other non-molecular based testing. Prevalence data was also collected after test implementation (03-30-16) through 04-30-16. Workflow metrics were documented and comparisons were made to show differences between the traditional in-house testing and the newer multiplex molecular testing. GPP testing is batched and run once per day for each day of the week.

Results: The GPP offered additional stool pathogen targets compared to the Microbiology laboratory offering of culture (*Campylobacter, Salmonella, Shiqella, E.* coli O157, and Yersinia), Shiga-toxin (EIA), and rotavirus antigen. The time to result for a sample after being received in the laboratory is approximately 6-7 hrs with the Medical Technologist manually entering the results. The Information Technology (IT) component of this test implementation was one of the bottlenecks in our process that resulted in going live with manual resulting and no interface. Our current laboratory information system (LIS) is SOFT Lab and our LIS vendor did not have any experience in setting up an interface with the Luminex Magpix instrument. We have a target turn-around-time (TAT) of 24 hrs. This 24-hr TAT is shorter when compared to our average stool culture TAT of 70.5 hrs. Additional send-out testing also takes multiple days to get results. GPP testing of the left-over stool samples showed additional detections for Norovirus that is normally not ordered or tested for routinely within our healthcare system. Prevalence data after test implementation showed Norovirus to be the most prevalent pathogen among pediatric and adult patients tested. Our Microbiology stool bench was discontinued allowing those FTEs to be devoted other responsibilities as well as being able to develop and implement new testing.

Conclusion: Implementation of the GPP within our healthcare system has given clinicians a wider breadth of GI pathogens than what we used to offer prior to molecular testing. The shorter TAT has allowed clinicians to act appropriately sooner based on the availability of results. The discontinuation of the stool bench in our Microbiology lab has increased our FTE resources to allocate to other responsibilities such as implementation of new testing and other bench-work that requires more hands-on time.

Materials and Methods

Samples

- Zeptometrix NATrol[™] samples
- Luminex provided samples
- Clinical diarrheal stool samples
- Raw and Cary-Blair diarrheal stool

Instruments

- easyMag Nucleic Acid Extractor (bioMerieux)
- Thermalcycler (Eppendorf)
- Luminex MAGPIX[®]

Reagents

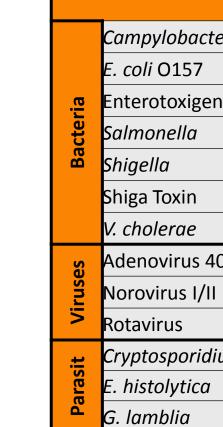
xTAG[®] GPP reagents. Assay performed as per manufacturer's protocol.

Data Analyses

- Data collection involved our IT Laboratory team and use of our LIS (SOFTLab)
- Data analyses were done using Microsoft Excel (Microscoft)



R	esul	ts — W	/orkflow/Metl	hodology Con	nparisons		
Method			Tests For	ТАТ	Positivity Rate (2014 volumes)		
Stool Cx			5 bacterial pathogens	48-72 hrs	12.5% (5,516)		
Ova & Parasite Exam			Parasite Eggs and Pathogens	24hrs	0.6% (1,664)		
Modified Acid-Fast Stain (for Cryptosporidium, Cyclospora, Isospora)		3 Parasites	24 hrs	4.2% (120)			
Rapid Rotav	Rapid Rotavirus Antigen		Rotavirus	~30 minutes	12.2% (377)		
			Luminex GPP				
	Campy						
		Сатру	lobacter				
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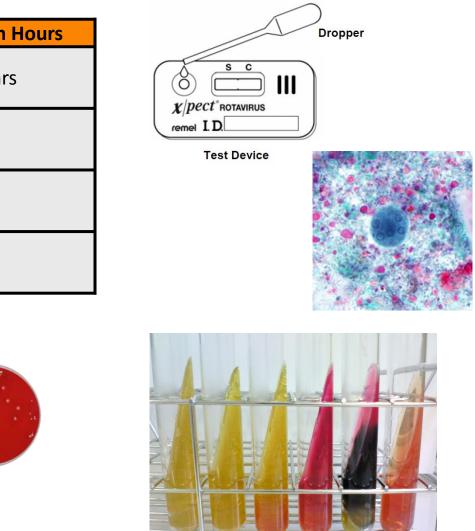


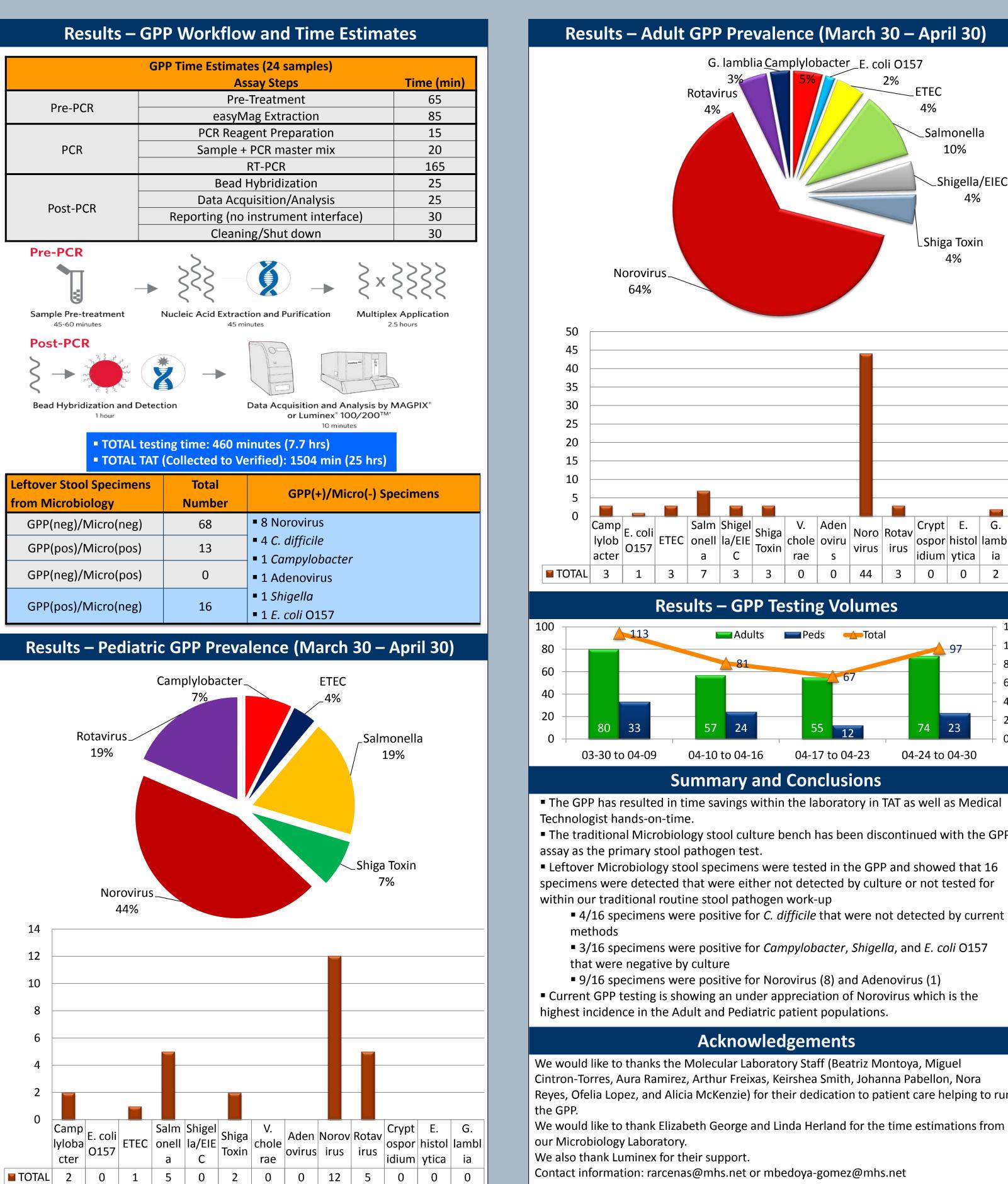
Results – Traditional In-house Testing Time Estimates

Stool Culture/O&P/Rotavirus Time Estimates					
		Technologist Time (min)			
	Receiving/processing	10 min			
Pre-Analytical	Stool Culture plating and other set-ups	Culture plating: 5 min			
Analytical	Stool Cx Identification	6-40 minutes (not including incubation time)			
	O&P	<60 minutes			
	Shiga Toxin EIA	<30 minutes			
	Rotavirus Antigen	18 minutes			
Post- Analytical	Manual Resulting	1-5 minutes			

Total Turn-Around-Time (TAT)				
Stool Cx Identification	48-72 ł			
O&P	<8 hrs			
Shiga Toxin EIA	<8 hrs			
Rotavirus Antigen	<1 hr			









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irusSporhistolIamblortarOOOOOOOOOO **TOTAL** 3 1 3 7 3 3 0 0 44 3 0 0 2 100 80 60 40 20

The GPP has resulted in time savings within the laboratory in TAT as well as Medical

The traditional Microbiology stool culture bench has been discontinued with the GPP

Leftover Microbiology stool specimens were tested in the GPP and showed that 16 specimens were detected that were either not detected by culture or not tested for

- 4/16 specimens were positive for C. difficile that were not detected by current
- 3/16 specimens were positive for Campylobacter, Shigella, and E. coli O157
- Current GPP testing is showing an under appreciation of Norovirus which is the

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