Development of a Semi-Quantitative Women's Health Bacterial Panel on the Luminex[®] NxTAG[®] Platform

<u>D. Irwin</u>¹, N. Pelte¹, L. Mazur², I. Chaplyk², M. Mihalov², H. Zhang¹ ¹Luminex Molecular Diagnostics Inc., Toronto, Canada, ²ACL Laboratories, Rosemont, Illinois, USA

Background and Objective

The women's health bacterial panel (WHBP) prototype in development highlights the semi-quantitative capabilities of the Luminex[®] NxTAG[®] platform. The WHBP prototype offers simplified and high throughput testing of 1-96 samples workflow of NxTAG (Figure 1) while simultaneously detecting and providing a semi-quantitative result for *Atopobium vaginae*, bacterial vaginosis associated bacteria 2 (BVAB2), and *Megasphaera* Type 1 (*Mega* 1).

The objective of this study was to evaluate the performance of the NxTAG WHBP panel on remnant gynecological (GYN) samples. High positive, low positive, and negative calls were determined for all analytes in the panel and realtime PCR (qPCR) was used as the comparator.



Step 1: Add extracted samples to wells containing pre-plated lyophilized reagents



Step 2: Reseal reaction vessel



Step 3: Transfer reaction vessel to thermal cycler



Step 4: Transfer reaction vessel to MAGPIX®

Figure 1. Overall Assay Workflow of NxTAG WHBP Prototype

Results

Table 1: Summary of NxTAG WHBP Results Confirmed by qPCR

A. vaginae Result	# of Calls	# qPCR Confirmed	% Confirmed	
High Positive	146	144	98.6	
Low Positive	12	12	100	
Negative	65	61	93.8	
Total	223	217	97.3	
BVAB2 Result	# of Calls	# qPCR Confirmed	% Confirmed	
High Positive	110	106	96.4	
Low Positive	15	12	80	
Negative	98	92	93.9	
Total	223	210	94.2	
<i>Mega</i> 1 Result	# of Calls	# qPCR Confirmed	% Confirmed	
High Positive	120	120	100	
Low Positive	43	30 69.8		
Negative	60	49 81.7		
Total	223	199	89.2	

A. vaginae: High positive > $3.2x10^5$ copies/ml; low positive = $3x10^2$ – $3.2x10^5$ copies/ml. **BVAB2**: High positive > $1.2 x10^5$ copies/ml; low positive = $3.2x10^2 - 1.2x10^5$ copies/ml. *Mega* 1: High positive > $1.2x10^5$ copies/ml; low positive = $5x10^2 - 1.2x10^5$ copies/ml

Table 2: Results of NxTAG WHBP Compared to Sample Nugent Score

Nugent Score	# of Samples with 2 or More High Positive Calls	# of Samples with 1 or Fewer High Positive Calls		
7-9	39	1		
4-6	27	3		

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Materials and Methods

A total of 223 remnant GYN samples were used for the study. Nucleic acids were extracted from 200 µl of raw sample collected in either PreservCyt, Universal Transport Media, or Liquid Amies using the NucliSENS[®] easyMAG[®] extractor and eluted in to 110 µl of elution buffer. An endogenous human control is used to eliminate the need for an external extraction control. Extracted nucleic acid was stored at -80°C until testing. 35 µl of extracted nucleic acid were added directly to preplated WHBP specific lyophilized bead reagents (LBRs) and NxTAG universal LBRs. Multiplexed PCR and bead hybridizations were performed in closed PCR tubes under a single cycling program. The sealed plates were placed directly on the MAGPIX[®] instrument for data acquisition. Thresholds applied to make calls are preliminary. Real-time PCR was used to confirm all calls for the three bacterial analytes detected by the NxTAG WHBP prototype.

Conclusion

- The NxTAG WHBP prototype is able to semi-quantitatively detect all three analytes in remnant GYN samples. 97.3%, 94.2%, and 89.2% of all NxTAG WHBP calls were confirmed by qPCR for *A. vaginae*, BVAB2, and *Mega* 1, respectively. The NxTAG WHBP gives a semi-quantitative result that compares well with the absolute quantification of qPCR.
- The NxTAG WHBP prototype results also correlate well with the Nugent Score (NS). 97.5% of samples with a NS = 7-9 and 90% of samples with NS= 4-6 were high positive for at least two analytes by the WHBP prototype.
- The NxTAG WHBP prototype in development offers a high throughput and scalable (1-96 samples) platform for the semiquantitative testing of three bacterial pathogens. The NxTAG platform also has the benefit to add additional analytes that require either a semi-quantitative or qualitative result.